

Diagnostic Sampling Protocol



The information provided below is for routine diagnostic investigation. For specific disease investigations call [TheFishVet](http://TheFishVet.com.au).

Sampling Fish

1. Choose representative fish
 - 3 pools:
 - i. Affected tank: 3-10 sick fish
 - ii. Affected tank: 3-10 healthy fish
 - iii. Unaffected tank: 3-10 healthy fish
2. Euthanase (please indicate method of euthanasia).
3. Midline incision and then aseptically swab kidney (Fig. 1).
4. Remove one operculum and coelomic wall (Fig. 2) to allow penetration of preservative. If fish are >10cm, then organs need to be collected individually.



Fig. 1



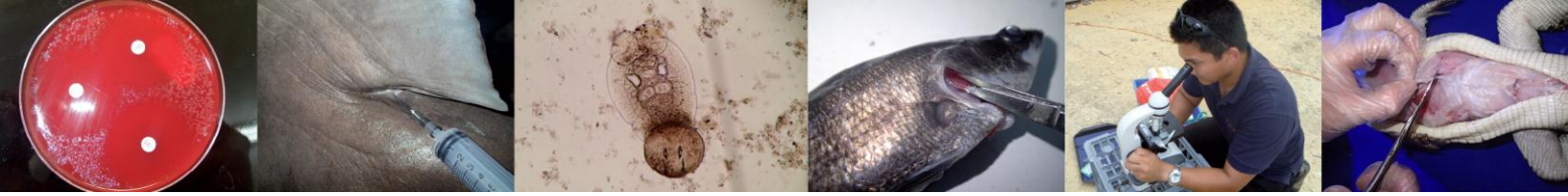
Fig. 2

5. Preserve fish in 10% NBF (Fig. 3).
6. Collect water samples.



Fig. 3





Packaging Samples

1. After 24h fixation at room temperature, the formalin from the containers may be drained.
2. Bag up the jars, and include absorbent paper to contain spillage.
3. Water samples, and bacteriology samples should be bagged separately (to avoid cross-contamination in the event of breakage).
4. Package swabs chilled, and package fixed-samples at room temperature.



Fig. 4

Additional Notes

- NBF Solution for preserving tissues is Neutral Buffered Formalin.
If none available, make up solution using 1 part formaldehyde solution (37%) to 9 parts fresh water.
- Swab for bacteriology contains Amies Charcoal Transport Medium.
- **Sampling equipment** can be ordered from [TheFishVet](http://TheFishVet.com.au).

If you have any questions please do not hesitate to contact me.

Yours sincerely,

Dr Richmond Loh

*DipProjMgt, BSc, BVMS, MPhil (Pathology) Murdoch,
MANZCVS (Aquatics & Pathobiology), CertAqV, NATA Sig.*

Aquatic Veterinarian & Veterinary Pathologist

Perth, Western Australia, AUSTRALIA.

